



AssayMax[™]

Human Factor V ELISA Kit

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For any questions regarding troubleshooting or performing the assay, please contact our support team at support@assaypro.com.

Thank you for choosing Assaypro.

Assay Summary

Step 1. Add 50 μ l of Standard or Sample per well.
Incubate 2 hours.

Step 2. Wash, then add 50 μ l of Biotinylated Antibody per well.
Incubate 1 hour.

Step 3. Wash, then add 50 μ l of SP Conjugate per well.
Incubate 30 minutes.

Step 4. Wash, then add 50 μ l of Chromogen Substrate per well.
Incubate 20 minutes.

Step 5. Add 50 μ l of Stop Solution per well.
Read at 450 nm immediately.

Symbol Key



Consult instructions for use.

Assay Template

[illegible]

Human Factor V ELISA Kit

Catalog No. EF1005-1

Sample protocol for reference use only

Introduction

Factor V (FV) is an essential cofactor of the blood coagulation cascade and circulates in plasma as a large single-chain glycoprotein. The deduced amino acid sequence consists of 2224 amino acids inclusive of a 28-amino acid leader peptide (1). During coagulation, it is converted to the active cofactor FVa via limited proteolysis by thrombin, and spliced into a heavy chain (110 KDa) and a light chain (73 KDa) held together non-covalently by calcium (2). In the presence of a calcium ion and the phospholipid on cell surfaces, FVa and FXa form the prothrombinase complex which catalyzes the hydrolysis of prothrombin to thrombin (3). Thrombin in turn cleaves fibrinogen to fibrin which polymerizes to form a clot. FVa is readily inactivated by anticoagulant activated protein C (4). FV Leiden, a single amino acid mutation, renders FVa resistant to cleavage by activated protein C. It therefore over-produces thrombin and leads to excess clotting and hereditary thrombophilia disorder (5). Severe FV deficiency is associated with mild to severe bleeding diathesis (6).

Principle of the Assay

The AssayMax Human Factor V ELISA (Enzyme-Linked Immunosorbent Assay) kit is designed for detection of human factor V in **plasma, urine, milk, CSF, and cell culture samples**. This assay employs a quantitative **sandwich enzyme immunoassay** technique that measures factor V in less than 4 hours. A monoclonal antibody specific for factor V has been pre-coated onto a 96-well microplate with removable strips. Factor V in standards and samples is sandwiched by the immobilized antibody and a biotinylated polyclonal antibody specific for factor V, which is recognized by a streptavidin-peroxidase conjugate. All unbound material is washed away and a peroxidase enzyme substrate is added. The color development is stopped and the intensity of the color is measured.

Caution and Warning

- Prepare all reagents (working diluent buffer, wash buffer, standard, biotinylated antibody, and SP conjugate) as instructed, prior to running the assay.

- Prepare all samples prior to running the assay. The dilution factors for the samples are suggested in this protocol. However, the user should determine the optimal dilution factor.
- Spin down the SP conjugate vial and the biotinylated antibody vial before opening and using contents.
- The Stop Solution is an acidic solution.
- This kit is for research use only.
- The kit should not be used beyond the expiration date.

Reagents

- **Human Factor V Microplate:** A 96-well polystyrene microplate (12 strips of 8 wells) coated with a monoclonal antibody against factor V.
- **Sealing Tapes:** Each kit contains 3 precut, pressure sensitive sealing tapes that can be cut to fit the format of the individual assay.
- **Human Factor V Standard:** Human factor V in a buffered protein base (210 ng, lyophilized).
- **Biotinylated Human Factor V Antibody (60x):** An 60-fold concentrated biotinylated polyclonal antibody against human factor V (100 µl).
- **EIA Diluent Concentrate (10x):** A 10-fold concentrated buffered protein base (30 ml).
- **Wash Buffer Concentrate (20x):** A 20-fold concentrated buffered surfactant (30 ml, 2 bottles).
- **Streptavidin-Peroxidase Conjugate (SP Conjugate):** A 100-fold concentrate (80 µl).
- **Chromogen Substrate:** A ready-to-use stabilized peroxidase chromogen substrate tetramethylbenzidine (8 ml).
- **Stop Solution:** A 0.5 N hydrochloric acid to stop the chromogen substrate reaction (12 ml).

Storage Condition

- Upon arrival, immediately store components of the kit at recommended temperatures up to the expiration date.
- Store SP Conjugate and Biotinylated Antibody at -20°C.
- Store Microplate, Diluent Concentrate (10x), Wash Buffer, Stop Solution, and Chromogen Substrate at 2-8°C.
- Unused microplate wells may be returned to the foil pouch with the desiccant packs and resealed. May be stored for up to 30 days in a vacuum desiccator.
- Diluent (1x) may be stored for up to 30 days at 2-8°C.
- Store Standard at 2-8°C before reconstituting with Diluent and at -20°C after reconstituting with Diluent

Other Supplies Required

- Microplate reader capable of measuring absorbance at 450 nm.
- Pipettes (1-20 µl, 20-200 µl, 200-1000 µl, and multiple channel).
- Deionized or distilled reagent grade water.

Sample Collection, Preparation, and Storage

- **Plasma:** Collect plasma using one-tenth volume of 0.1 M sodium citrate as an anticoagulant. Centrifuge samples at 3000 x *g* for 10 minutes. Dilute samples 1:800 with EIA Diluent and assay. The undiluted samples can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles (EDTA or Heparin can also be used as an anticoagulant).
- **Cell Culture Supernatants:** Centrifuge cell culture media at 3000 x *g* for 10 minutes to remove debris. Collect supernatants and assay. Store the remaining samples at -20°C or below. Avoid repeated freeze-thaw cycles.
- **Urine:** Collect urine using sample pot. Centrifuge samples at 800 x *g* for 10 minutes. Dilute samples 1:2 with EIA Diluent and assay. The undiluted samples can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.
- **Milk:** Collect milk using sample tube. Centrifuge samples at 800 x *g* for 10 minutes. Dilute samples 1:2 with EIA Diluent and assay. The undiluted samples can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.
- **CSF:** Collect cerebrospinal fluid (CSF) using sample pot. Centrifuge samples at 3000 x *g* for 10 minutes. Dilute samples 1:2 into EIA Diluent and assay. The undiluted samples can be stored at -80°C for up to 3 months. Avoid repeated freeze-thaw cycles.

Refer to Sample Dilution Guidelines below for further instruction.

Guidelines for Dilutions of 1:100 or Greater <i>(for reference only; please follow the protocol for specific dilution suggested)</i>	
1:100	1:10000
A) 4 µl sample : 396 µl buffer (100x) = 100 fold dilution <i>Assuming the needed volume is less than or equal to 400 µl.</i>	A) 4 µl sample : 396 µl buffer (100x) B) 4 µl of A : 396 µl buffer (100x) = 10000 fold dilution <i>Assuming the needed volume is less than or equal to 400 µl.</i>
1:1000	1:100000
A) 4 µl sample : 396 µl buffer (100x) B) 24 µl of A : 216 µl buffer (10x) = 1000 fold dilution <i>Assuming the needed volume is less than or equal to 240 µl.</i>	A) 4 µl sample : 396 µl buffer (100x) B) 4 µl of A : 396 µl buffer (100x) C) 24 µl of B : 216 µl buffer (10x) = 100000 fold dilution <i>Assuming the needed volume is less than or equal to 240 µl.</i>

Reagent Preparation

- Freshly dilute all reagents and bring all reagents to room temperature before use.
- EIA Diluent Concentrate (10x):** If crystals have formed in the concentrate, mix gently until the crystals have completely dissolved. Dilute EIA Diluent Concentrate 1:10 with reagent grade water. Store for up to 30 days at 2-8°C.
- Standard Curve:** Reconstitute the 210 ng of Human Factor V Standard with 3.5 ml of EIA Diluent to generate a 60 ng/ml standard stock solution. Allow the standard to sit for 10 minutes with gentle agitation prior to making dilutions. Prepare duplicate or triplicate standard points by serially diluting the standard stock solution (60 ng/ml) 1:2 with EIA Diluent to produce 30, 15, 7.5, 3.75, 1.875, and 0.938 ng/ml solutions. EIA Diluent serves as the zero standard (0 ng/ml). Any remaining solution should be frozen at -20°C and used within 30 days.

Standard Point	Dilution	[FV] (ng/ml)
P1	1 part Standard (60 ng/ml)	60.00
P2	1 part P1 + 1 part EIA Diluent	30.00
P3	1 part P2 + 1 part EIA Diluent	15.00
P4	1 part P3 + 1 part EIA Diluent	7.500
P5	1 part P4 + 1 part EIA Diluent	3.750
P6	1 part P5 + 1 part EIA Diluent	1.875
P7	1 part P6 + 1 part EIA Diluent	0.938
P8	EIA Diluent	0.000

- **Biotinylated Human Factor V Antibody (60x):** Spin down the antibody briefly and dilute the desired amount of the antibody 1:60 with EIA Diluent. Any remaining solution should be frozen at -20°C.
- **Wash Buffer Concentrate (20x):** If crystals have formed in the concentrate, mix gently until the crystals have completely dissolved. Dilute Wash Buffer Concentrate 1:20 with reagent grade water.
- **SP Conjugate (100x):** Spin down the SP Conjugate briefly and dilute the desired amount of the conjugate 1:100 with EIA Diluent. Any remaining solution should be frozen at -20°C.

Assay Procedure

- Prepare all reagents, standard solutions, and samples as instructed. Bring all reagents to room temperature before use. The assay is performed at room temperature (20-25°C).
- Remove excess microplate strips from the plate frame and return them immediately to the foil pouch with desiccants inside. Reseal the pouch securely to minimize exposure to water vapor and store in a vacuum desiccator.
- Add 50 µl of Human Factor V Standard or sample per well. Cover wells and incubate for 2 hours. Start the timer after the last addition.
- Wash five times with 200 µl of Wash Buffer manually. Invert the plate each time and decant the contents; hit 4-5 times on absorbent material to completely remove the liquid. If using a machine, wash six times with 300 µl of Wash Buffer and then invert the plate, decanting the contents; hit 4-5 times on absorbent material to completely remove the liquid.
- Add 50 µl of Biotinylated Human Factor V Antibody to each well and incubate for 1 hour.
- Wash the microplate as described above.
- Add 50 µl of Streptavidin-Peroxidase Conjugate per well and incubate for 30 minutes. Turn on the microplate reader and set up the program in advance.
- Wash the microplate as described above.
- Add 50 µl of Chromogen Substrate per well and incubate for 20 minutes or till the optimal blue color density develops. Gently tap the plate to ensure thorough mixing and break the bubbles in the well with pipette tip.
- Add 50 µl of Stop Solution to each well. The color will change from blue to yellow.
- Read the absorbance on a microplate reader at a wavelength of 450 nm **immediately**. If wavelength correction is available, subtract readings at 570 nm from those at 450 nm to correct optical imperfections. Otherwise, read the plate at 450 nm only. Please note that some unstable black particles may be generated at high concentration points

after stopping the reaction for about 10 minutes, which will reduce the readings.

Data Analysis

- Calculate the mean value of the duplicate or triplicate readings for each standard and sample.
- To generate a standard curve, plot the graph using the standard concentrations on the x-axis and the corresponding mean 450 nm absorbance (OD) on the y-axis. The best-fit line can be determined by regression analysis using log-log or four-parameter logistic curve-fit.
- Determine the unknown sample concentration from the Standard Curve and multiply the value by the dilution factor.

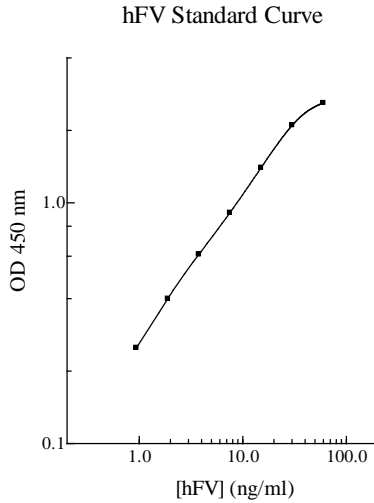
Typical Data

- The typical data is provided for reference only. Individual laboratory means may vary from the values listed. Variations between laboratories may be caused by technique differences.

Standard Point	ng/ml	OD	Average OD
P1	60.00	1.857 1.816	1.836
P2	30.00	1.504 1.502	1.503
P3	15.00	1.123 1.138	1.131
P4	7.500	0.788 0.802	0.795
P5	3.750	0.550 0.599	0.574
P6	1.875	0.490 0.500	0.495
P7	0.938	0.371 0.375	0.373
P8	0.000	0.213 0.209	0.211
Sample: Pool Normal, Sodium Citrate Plasma (800x)		0.894 0.898	0.896

Standard Curve

- The curve is provided for illustration only. A standard curve should be generated each time the assay is performed.



Reference Values

- Normal human factor V plasma levels range from 4 to 16 $\mu\text{g/ml}$.
- Human plasma samples from healthy adults were tested ($n=20$). On average, factor V level was 8.8 $\mu\text{g/ml}$.

Performance Characteristics

- The minimum detectable dose of factor V as calculated by 2SD from the mean of a zero standard was established to be 0.9 ng/ml.
- Intra-assay precision was determined by testing replicates of three plasma samples in one assay.
- Inter-assay precision was determined by testing three plasma samples in twenty assays.

	Intra-Assay Precision			Inter-Assay Precision		
Sample	1	2	3	1	2	3
n	20	20	20	20	20	20
CV (%)	4.9%	5.1%	4.8%	9.9%	9.7%	8.8%
Average CV (%)	4.9%			9.5%		

Spiking Recovery

- Recovery was determined by spiking two plasma samples with different factor V concentrations.

Sample	Unspiked Sample (ng/ml)	Spike (ng/ml)	Expected	Observed	Recovery (%)
1	5.2	2.5	7.7	8.1	105%
		5.0	10.2	10.3	101%
		10.0	15.2	14.9	98%
2	2.4	2.5	4.9	5.4	110%
		5.0	7.4	7.5	101%
		10.0	12.4	11.9	96%
Average Recovery (%)					102%

Linearity

- Plasma samples were serially-diluted to test for linearity.

Average Percentage of Expected Value (%)	
Sample Dilution	Plasma
1:400	94%
1:800	98%
1:1600	105%

Cross-Reactivity

Species	Cross Reactivity (%)
Canine	None
Bovine	None
Monkey	<15%
Mouse	None
Rat	None
Swine	<15%
Rabbit	None

Troubleshooting

Issue	Causes	Course of Action
Low Precision	Use of expired components	<ul style="list-style-type: none"> Check the expiration date listed before use. Do not interchange components from different lots.
	Improper wash step	<ul style="list-style-type: none"> Check that the correct wash buffer is being used. Check that all wells are dry after aspiration. Check that the microplate washer is dispensing properly. If washing by pipette, check for proper pipetting technique.
	Splashing of reagents while loading wells	<ul style="list-style-type: none"> Pipette properly in a controlled and careful manner.

Unexpectedly Low or High Signal Intensity	Inconsistent volumes loaded into wells	<ul style="list-style-type: none"> Pipette properly in a controlled and careful manner. Check pipette calibration. Check pipette for proper performance.
	Insufficient mixing of reagent dilutions	<ul style="list-style-type: none"> Thoroughly agitate the lyophilized components after reconstitution. Thoroughly mix dilutions.
	Improperly sealed microplate	<ul style="list-style-type: none"> Check the microplate pouch for proper sealing. Check that the microplate pouch has no punctures. Check that three desiccants are inside the microplate pouch prior to sealing.
	Microplate was left unattended between steps	<ul style="list-style-type: none"> Each step of the procedure should be performed uninterrupted.
	Omission of step	<ul style="list-style-type: none"> Consult the provided procedure for complete list of steps.
	Steps performed in incorrect order	<ul style="list-style-type: none"> Consult the provided procedure for the correct order.
	Insufficient amount of reagents added to wells	<ul style="list-style-type: none"> Check pipette calibration. Check pipette for proper performance.
	Wash step was skipped	<ul style="list-style-type: none"> Consult the provided procedure for all wash steps.
	Improper wash buffer	<ul style="list-style-type: none"> Check that the correct wash buffer is being used.
	Improper reagent preparation	<ul style="list-style-type: none"> Consult reagent preparation section for the correct dilutions of all reagents.
Deficient Standard Curve Fit	Insufficient or prolonged incubation periods	<ul style="list-style-type: none"> Consult the provided procedure for correct incubation time.
	Non-optimal sample dilution	<ul style="list-style-type: none"> Sandwich ELISA: If samples generate OD values higher than the highest standard point (P1), dilute samples further and repeat the assay. Competitive ELISA: If samples generate OD values lower than the highest standard point (P1), dilute samples further and repeat the assay. User should determine the optimal dilution factor for samples.
	Contamination of reagents	<ul style="list-style-type: none"> A new tip must be used for each addition of different samples or reagents during the assay procedure.
	Contents of wells evaporate	<ul style="list-style-type: none"> Verify that the sealing film is firmly in place before placing the assay in the incubator or at room temperature.
	Improper pipetting	<ul style="list-style-type: none"> Pipette properly in a controlled and careful manner. Check pipette calibration. Check pipette for proper performance.
	Insufficient mixing of reagent dilutions	<ul style="list-style-type: none"> Thoroughly agitate the lyophilized components after reconstitution. Thoroughly mix dilutions.

References

- (1) Jenny RJ *et al.* (1987) Proc. Natl. Acad. Sci. U.S.A. 84:4846-4850
- (2) Esmon CT (1979) J. Biol. Chem. 254:964-973
- (3) Nesheim ME *et al.* (1981) J. Biol. Chem. 256:6537-6540
- (4) Kisiel W *et al.* (1977) Biochemistry 16:5824-5831
- (5) Bertina RM *et al.* (1994) Nature 369:64-67
- (6) Asselta R *et al.* (2006) J. Thromb. Haemost 4(1):26-34

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